Supplementary Online Content


**eFigure 1.** Puerto Rico Department of Health Guillain-Barré Syndrome Case Report Form

**eFigure 2.** Case Identification Flow Diagram

**eTable 1.** Zika Virus Diagnostic Test Results Among Confirmed GBS Cases With Evidence of Zika Virus Infection (N = 71)

**eTable 2.** Detection of Zika Virus RNA and Anti-Zika Virus IgM Antibodies Among Guillain-Barré Syndrome Patients’ Body Fluids (Serum, Urine, and Cerebrospinal Fluid) by Day Prior to and Post Neurologic Illness Onset, Puerto Rico, 2016

This supplementary material has been provided by the authors to give readers additional information about their work.
### eFigure 1. Puerto Rico Department of Health Guillain-Barré Syndrome Case Report Form

The health provider (neurologist/physician) will complete in print the GBS Surveillance Report Form and will mark the laboratory test abnormal panel (elevated IgM, elevated IgG, and AChR). The laboratory will submit filled out form and serum sample. Additional samples (e.g., urine, saliva, and cerebrospinal fluid) can be submitted in addition to form and serum sample.

**Instructions to fill the Guillain-Barré Syndrome (GBS) Surveillance Report Form**

- **Sections I and II.** The patient’s complete name and information is essential because many persons have similar names and information. Complete as instructed.
- **Section III.** Complete no other symptoms. A list of diagnostic manifestations and not antecedent symptoms. Please report any first encounter with a healthcare provider (present or out-patient).
- **GBS diagnosis.** The date of diagnosis of GBS is rendered by a neurologist or physician.
- **Other symptoms.** Please check all manifestations noted until date of admission of case report form (present at any time during illness).
- **Sections IV, VI.** Please report any antecedent symptoms/conditions present at the time and any diseases of the case.

*Please fill out any information available at the time of report submission, but do not delay reporting in order to complete all sections. Please make a copy of this form. Complete additional information and submit all information at patient’s discharge. Subsequent serum samples can be tested using the original panel and an updated filled out form.*

**Available online:**


© 2018 American Medical Association. All rights reserved.
**eFigure 2. Case Identification Flow Diagram**

Cases identified by passive reporting
N = 135

Cases identified by diagnostic code
N = 181
- Already reported: n = 100
- Other final diagnosis: n = 6
- Hospitalized <3 days: n = 3

Total individual cases followed-up by chart review
n = 207

Confirmed GBS cases
n = 123
- With evidence of ZIKV
  n = 71
  - RT-PCR: n = 28
  - ZIKV & DENV/IM ELISA: n = 25
- Without evidence of ZIKV
  n = 36
  - RT-PCR: n = 18
  - ZIKV & DENV/IM ELISA: n = 18
- No specimens
  n = 16

Suspected GBS cases
n = 12
- With evidence of ZIKV
  n = 5
  - RT-PCR: n = 1
  - ZIKV & DENV/IM ELISA: n = 1
- Without evidence of ZIKV
  n = 5
- No specimens
  n = 2

Non-cases
n = 72
**eTable 1. Zika Virus Diagnostic Test Results Among Confirmed GBS Cases With Evidence of Zika Virus Infection (N = 71)**

<table>
<thead>
<tr>
<th>Test Description</th>
<th>n (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Zika virus nucleic acid detected by rRT-PCR</td>
<td>28 (39)</td>
</tr>
<tr>
<td>Anti-Zika virus IgM antibodies detected by ELISA</td>
<td>25 (35)</td>
</tr>
<tr>
<td>Anti-Zika virus and anti-dengue virus IgM antibodies detected by ELISA</td>
<td>18 (25)</td>
</tr>
</tbody>
</table>

ELISA = Enzyme-linked immunosorbent assay; IgM = Immunoglobulin M; rRT-PCR = Real-time reverse transcriptase-polymerase chain reaction
**eTable 2.** Detection of Zika Virus RNA and Anti-Zika Virus IgM Antibodies Among Guillain-Barré Syndrome Patients’ Body Fluids (Serum, Urine, and Cerebrospinal Fluid) by Day Prior to and Post Neurologic Illness Onset, Puerto Rico, 2016

<table>
<thead>
<tr>
<th>Days Prior to Neurologic Illness Onset</th>
<th>Serum</th>
<th>Urine</th>
<th>CSF</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>rRT-PCR</td>
<td>Anti-ZIKV IgM ELISA</td>
<td>rRT-PCR</td>
</tr>
<tr>
<td>N = 99</td>
<td>N = 95</td>
<td>N = 43</td>
<td>N = 31</td>
</tr>
</tbody>
</table>

### Number/Total Number of Specimens (%)

<table>
<thead>
<tr>
<th>Days Post Neurologic Illness Onset</th>
<th>Serum</th>
<th>Urine</th>
<th>CSF</th>
</tr>
</thead>
<tbody>
<tr>
<td>0–4</td>
<td>9/29 (31)</td>
<td>27/28 (96)</td>
<td>1/9 (11)</td>
</tr>
<tr>
<td>5–9</td>
<td>8/36 (22)</td>
<td>33/33 (100)</td>
<td>4/18 (22)</td>
</tr>
<tr>
<td>10–14</td>
<td>1/13 (8)</td>
<td>15/16 (94)</td>
<td>1/8 (13)</td>
</tr>
<tr>
<td>15–19</td>
<td>1/4 (25)</td>
<td>5/5 (100)</td>
<td>0/1 (0)</td>
</tr>
<tr>
<td>20–24</td>
<td>0/2 (0)</td>
<td>2/2 (100)</td>
<td>0/1 (0)</td>
</tr>
<tr>
<td>25–29</td>
<td>2/3 (67)</td>
<td>2/3 (67)</td>
<td>0/3 (0)</td>
</tr>
<tr>
<td>30–34</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>35–39</td>
<td>1/1 (100)</td>
<td>1/1 (100)</td>
<td>0/1 (0)</td>
</tr>
<tr>
<td>40–44</td>
<td>—</td>
<td>—</td>
<td>—</td>
</tr>
<tr>
<td>≥45</td>
<td>0/3 (0)</td>
<td>2/3 (67)</td>
<td>0/2 (0)</td>
</tr>
</tbody>
</table>

CSF = cerebrospinal fluid; ELISA = Enzyme-linked immunosorbent assay; IgM = Immunoglobulin M; rRT-PCR = Real-time reverse transcriptase-polymerase chain reaction; ZIKV = Zika virus.

— indicate that no specimens from GBS patients with evidence of ZIKV infection were collected during these date ranges.